Factors Affecting Chloride Uptake and Implications of the Chloride-Nitrate Antagonism in Sugarbeet Mineral Nutrition

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Chloride has been identified as an essential plant nutrient element for sugarbeets and other plants (7,11). Ulrich and Ohki (11) reported that C1 concentrations in sugarbeet petioles of 0.2 milliequivalents per gram (meq/g) were associated with healthy plants but that C1 concentrations approximating 0.005 meq/g were indicative of extreme C1 deficiency and growth retardation.

When CI is readily available the amounts absorbed by plants greatly exceed the growth-limiting levels. Kretchmer *et al.* (9) state that increasing levels of CI in the substrate resulted in linear increases in CI content of 10 different plant species. This effect was independent of type of substrate or plant species. They reported dry-weight CI concentrations up to about 1.2 meq/g of plant tissue.

Another aspect of C1 nutrition is that increasing levels of C1 in plants are associated with decreasing levels of NO₃-N and vice versa (1,2,6,9,10,12,13). Thus, there is an antagonism or negative interaction between these two ions in nutrient absorption.

These kinds of observations prompted an investigation into the role of C1 in central Washington-grown sugarbeets. Field experiments have been under way for several years which were not initially designed with the study of C1 in mind. C1 analyses proved to be very useful, however, in explaining some apparently anomalous results. In an earlier report (5), for example, it was shown that increasing rates of K fertilizer (as KC1) resulted in increasing levels of sucrose in the beet roots. Also, 'NO₈ in the petioles decreased with the K rates. The analytical results for petiole C1 (not reported earlier) are presented here in Figure 1. The results for K uptake are repeated for comparison. Figure 1 indicates that C1 concentration increased from below 0.2 meq/g to more than 1.2 meq/g. There is no indication in these results of an upper limit of C1 absorption. It is apparent also that K and C1 uptake are independent of each other.

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³ Numbers in parentheses refer to literature cited.

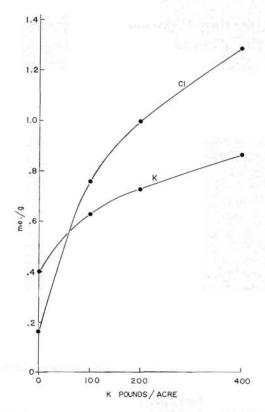


Figure 1.—Effect of applied K (as KC1) on K and C1 content of sugarbeet petioles. Each point is the mean of 12 replications. For further details see (5).

The basic objective of a series of experiments conducted in 1967 and 1968 was to evaluate sugarbeet responses to N, P and K fertilizers under central Washington conditions. A preliminary report on the N aspects of this research has been made (4). The purpose of this report is to present the results of Cl analyses from these experiments. Factors affecting Cl availability and the implications of the Cl-NO₃ interaction are discussed.

Procedure

Sugarbeets were studied at 40 different locations in 2 years. Data obtained included soil and petiole analyses and yield results. Experiments at 26 locations were carried to completion. Some information was recovered from several locations which were not carried to completion. Each experimental site consisted of approximately 1 acre in a larger commercial field of sugarbeets. Fields were selected which had not received any fertilizer

since the beginning of the previous growing season. Before crop establishment, the soil was sampled according to a standard procedure which takes into account the wide horizontal and vertical variations in soluble soil constituents which commonly occur in irrigated soils (3). Each experimental area was separated into 0.034 acre size plots and fertilizer treatments were applied according to the schedule given in Table 1. Phosphate fertilizer was used where necessary so that P was not a limiting factor in the growth of beets. In most instances the fertilizer was applied broadcast-plowdown. In a few cases the fertilizer was applied as sidedress after thinning. The fertilizer treatments were organized in a randomized complete block design in four replications. Planting and all cultural practices were done by the grower-cooperator.

Table 1.—Treatment combinations of nitrogen, and potassium fertilizer.

Treatment	Element rate—pounds per acre1		
No.	N	Almark Terri	\mathbb{K}^2
1	0		0
2	100		0
3	200		0
4	200		200

¹ The element forms were: N, ammonium nitrate, and K, muriate of potash.

² When the preliminary soil test indicated a probable K deficiency, treatments 1, 2 and 3 received K, and it was then omitted in treatment 4. Thus, treatment 4 was either a positive or negative check treatment, depending on the location.

The sampling and NO₃ analytical procedures for petioles have been described (5).

C1 was measured in distilled water extracts of soil and plant samples by an adaptation of the method given by Johnson and Ulrich (8). Petiole NO₃ and C1 are reported in terms of meq/g dry weight. Soil NO₃ and C1 are reported in terms of soil test indexes. Each index is the sum of the element ppm in all one-foot increments of soil through the zone sampled.

Results and Discussion

Petiole CI versus Soil Test Cl

Petiole Cl concentrations from the zero-K treatment at the respective experiments are presented in Figure 2 as a function of soil test Cl. The results follow the same pattern for both years. In general, as soil test Cl goes up the Cl content of petiole goes up. The (o) data points are from an area that is irrigated by return-flow water. This water contains 18 ppm of Cl. Seasonal fluctuations in this Cl load are quite small. All other data points are from experiments that are irrigated with water directly from the Yakima and Columbia rivers. These waters have Cl concentrations of 2.0 and 0.6 ppm, respectively.

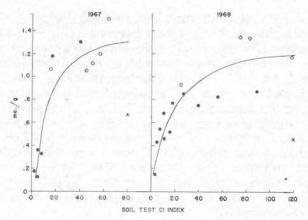


Figure 2.—The relationship between Cl concentration of sugarbeet petioles and the soil test Cl index is 2 years. Each point is the mean of four replications of treatment No. 1 from different experiments. See text for explanation of different symbols.

Some of the fields receiving low-C1 water have a history of KC1 fertilization. A few fields have undergone some degree of salinization.

The (x) data points are from fields that had a high amount of residual fertilizer N. Estimations from soil analyses indicate that the 1967 high-N field contained 310 pounds of NO₃-N per acre in the rooting zone, and the 1968 high-N field contained 460 pounds per acre. (These conversions are based on an average soil bulk density value of 1.45.) The results from these two sites indicate the depression of C1 uptake by NO₃.

It is apparent from Figure 2 that water-soluble C1 in the soil is a good indicator of C1 availability. It is apparent also that C1 availability at a given site can be predicted only after considering irrigation water input and the background of residual NO₃. The latter point will become more evident with subsequent discussions.

A very pertinent question arises from Figures 1 and 2. This has to do with the likelihood of having CI-deficient sugarbeets where both soil and water are very low in CI content. This question is being pursued.

The C1-NO₃ Negative Interaction

It has been shown that the soil test N index is a fair indicator of N availability for sugarbeets in central Washington (4). In light of this and the foregoing data on Cl uptake, the background on which the fertilizer treatments were superimposed should be considered in interpreting the treatment effects. Sugarbeet re-

sponses to the fertilizer treatments, in terms of petiole C1 and NO₃ concentrations, are given in Figures 3, 4 and 5 for seven selected sites. The soil test indexes for these sites are given in Table 2. The selections were made to show the different ways in which the C1-NO₃ uptake interaction manifested itself.

Table 2.-Soil test indexes for NOa-N and CI1 (background data for Figures 3, 4 and 5).

Soil test index				Figure			
	3 <i>a</i>	36	4a	4b	4c	5 <i>a</i>	56
N	36	69	8	26	13	13	78
CI	62	17	6	29	2	50	80

¹ Soil test index is the sum of the element ppm in all one-foot increments sampled, Sampling depths varied from 14 to 72 inches, depending on the depth of the rooting zone. The limiting soil layer was caliche.

Figure 3 shows two sets of results where 200 pounds of K per acre were applied with treatments 1, 2 and 3 (Table 1). The K rate was omitted with treatment 4. In Figure 3, as the N rate goes up the petiole NO₃ goes up and petiole Cl goes down. With treatment 4, Cl content drops and NO₃ increases. Note that without added KCl, there is more petiole Cl than NO₃ in Figure 3a and less Cl than NO₃ in Figure 3b. This shift is in agreement with the soil test indexes, Table 2.

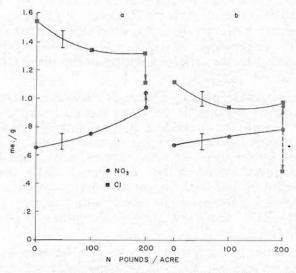


Figure 3.—The effect of $\mathrm{NH_4NO_3}$ and KCl fertilizer on $\mathrm{NO_3}$ and Cl concentrations of sugarbeet petioles. Each point is the mean of four replications. The vertical line segment is LSD (.05). Treatments 1, 2 and 3 included KCl at 200 pounds per acre of K. The effects of omitting KCl at treatment 4 are indicated by the arrows. Petioles were sampled (a) 6/22/67 and (b) 6/28/67.

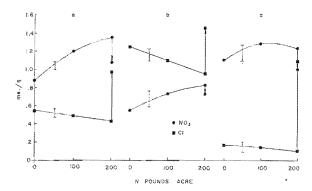


Figure 4.—Effect of NH₄NO₉ and KC1 fertilizer on NO₉ and C1 concentrations in sugarbeet petioles. KCl fertilizer was introduced only with treatment 4. Petioles were sampled (a) 6/18/68, (b) 6/21/68, and (c) 6/4/68.

In Figure 4 results are shown for three sites where KC1 was introduced only with treatment 4. At each site as the N fertilizer rate increased petiole NO $_3$ increased and with treatment 4 petiole NO $_3$ decreased. Petiole C1 decreased regularly with N rates in Figures 4a and b and increased upon addition of KC1. The level of soil C1 at the site represented by Figure 4c was apparently too low for the N rate to have much effect on C1 uptake. When KC1 was added, however, there was a very large effect on petiole C1. Again, the variation in soil test C1 is reflected in the results of Figure 4 by the relative position of the three C1 response curves.

At the site represented by Figure 5a there was a large petiole NO_3 response to N rates. In 5b there was no response to the N fertilizer. The effects of residual N (soil test 13 versus 78) are especially notable in these two sets of results. Petiole NO_3 was not affected by fertilizer KCl in Figure 5a; this kind of null response occurred infrequently. The petiole NO_3 response to KCl in Figure 5b followed the more common pattern. In both Figure 5a and b petiole Cl decreased with fertilizer N and increased with KCl. Only one possible exception to this pattern has been noted.

Another kind of contrast in results may be seen by comparing Figures 4c and 5a. In both of these cases soil test N was identical and low. The two experiments were in different years, but the shift in positions of petiole NO_5 and Cl is evidence of the different between Cl soil tests: Figure 4c had very low soil Cl background (coupled with low water Cl content); and Figure 5a had a relatively high soil Cl (as a result of water-borne Cl).

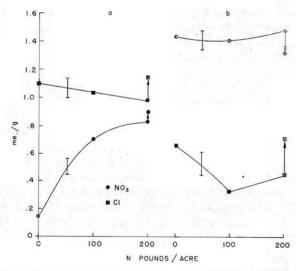


Figure 5.—Effects of NH_4NO_3 and KC1 fertilizer on NO_3 and C1 concentrations of sugarbeet petioles. KCl was introduced only with treatment 5. Petioles were sampled (a) 6/22/67 and (b) 6/20/67.

The C1-NO₃ negative interaction in plant nutrition has been investigated under solution or sand culture conditions (1,2,9, 10,12,13). From the results presented here it appears that the antagonism clearly manifests itself under field conditions too. It is apparent also that this phenomenon has important implications in assessing either soil test N as an index of N availability or petiole NO₃ as an indicator of the N nutritional status of the plant. From all the results taken from these experiments, for example, as a first approximation, optimum N fertilizer rates will be 50 pounds per acre higher in the areas irrigated with 18 ppm C1 waters compared to the areas receiving low-C1 waters.

Root Yield and Sucrose Responses

It is not the main purpose of this report to elaborate on the overall crop performance in this series of experiments. It is well to point out in passing, however, that there was one site where there was a possibility of a direct response to K per se as judged by petiole K and Na levels and root yields. In addition there were six sites that had a root yield response to 100 pounds of N per acre. (Among the sites represented by the figures there were significant responses to N in 4a, 4c and 5a.) In all cases there was a reduction in sucrose percent with added N fertilizer. The negative response of sucrose percent to increased N availability was quite sensitive.

Of direct concern here is the effect of the Cl-NO3 interaction on crop performance. In this respect, there was no detectable effect on root growth. There was, however, some effect on sucrose. It was pointed out previously (5) that positive sucrose responses to KCl fertilization have been noted quite frequently in this area. The increases ranged up to about 0.8%, generally averaging around 0.4%. Similar kinds of responses were observed in the 1967-68 experiments. These are summarized in Table 3. The criteria for judging the KCI effect was a difference of 0.2% sucrose between treatments 3 and 5 at each site. (The relative experimental error for sucrose percent is typically lower than for other plant nutrition and growth parameters. Differences of 0.2% sucrose are usually significant at the 1% level.) On this basis 54% of all experiments had a significant increase in sucrose, with 38% showing no effect. Eight percent showed a signficantly lower level of sucrose.

Table 3.—Changes in sucrose percent associated with additions of KCl fertilizer: A frequency distribution.

Type of response	No. of sites	Percent in category
Null	10	38
0.2% or greater increase	14	54
0.2% decrease	2	8
		-
Total	26	100

The nature of the sucrose response to KC1 fertilizer was not readily predicted by soil test or by tissue analyses. Nevertheless, when there was a positive sucrose response to KC1 fertilization, the most logical explanation is that the sugarbeet is responding to reduced NO₈ uptake. In other words, the change in sucrose is an indirect response to the C1-containing fertilizer. Many of the other factors that affect sucrose accumulation can be related to N. For example, sugarbeets under furrow irrigation typically show a reduction in sucrose after a rainstorm which was preceded by a prolonged period of no precipitation. In this instance, NO3-N has been washed from its position of isolation in the dry surface soil into the active root zone. On the other hand, the crop responds with increased content of sucrose to excessive irrigation as a result of N leaching. Thus, several unrelated variables could cancel the apparent effect of KCl on sucrose accumulation.

The foregoing interpretation of the plant C1-sucrose relationships assumes that there are no direct connections between K uptake and sucrose accumulation beyond the point where K content is non-limiting to normal metabolism. In the examples

given, the experimental design does not allow for a clear-cut separation of the effects of K and C1. But increases in percent sucrose between treatments 3 and 4 (N_{200} versus N_{200} K₂₀₀) occurred when K in the petiole ranged from .55 and 2.14 meg/g. These numbers are well above the tissue-K levels associated with K deficiency (5).

Summary and Conclusions

Experiments involving N and K fertilizers were conducted in commercial sugarbeet fields in 1967 and 1968. These experiments were not designed initially with C1 nutrition in mind but analyses of soils, irrigation waters, and plant materials were made in an attempt to explain some apparently anomalous results.

The investigations on CI indicate that Cl is readily absorbed by sugarbeets. Uptake of Cl is essentially a exponential function of the amount of water-soluble Cl in the soil. Cl in the plant was also related to the load of Cl in irrigation water. The data indicate that certain areas of central Washington have very low Cl concentrations in both soil and water. This raises a question on the possibility of Cl-deficient sugarbeets in the area.

Results from fertilization with NH_4NO_8 and KC1 confirm the well-documented antagonism in plant uptake of NO_8 and C1. The way in which the antagonism expresses itself can be related to the background concentrations of NO_9 and C1 in the soil. The evidence points up the necessity of analyzing soils and plants for both C1 and NO_8 in order to properly evaluate soil N availability and the N nutritional status of the plant.

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