Predicting Nitrogen Fertilizer Needs for Sugarbeets from Residual Nitrate and Mineralizable Nitrogen¹

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Nitrogen (N) fertilizer management for sugarbeet (*Beta vulgaris* L.) production requires more precise information than for most crops. Inadequate N limits plant growth and root yield, but excess N may reduce both sucrose percentage and recoverable sucrose (7).³ Also, excess N may stimulate more leaf growth than necessary. The rate and timing of N fertilizer applications are not only important in supplying crop N needs, but can influence the amount of N lost by leaching and denitrification. Soil and plant tissue tests can provide essential data for decision-making for efficient and economical use of N fertilizer.

Recent studies have shown that the NO₃-N level in the soil before planting is closely related to sucrose production when N is limiting (8, 12). Inclusion of the N mineralization capacity of the soils would be expected to improve the relationship. Stanford and Smith (14) showed that the mineralization capacity varies with soil type and location. Therefore, a soil test for N that would have general applicability should include the mineralization capacity of the soil, and the interpretation of these tests should include some knowledge of expected irrigation practices. A soil test for NO₃-N may suffice as an index of N fertilizer needs for a given soil and irrigation level.

Recently, Carter et al. (5) showed that sucrose production was closely related to available soil N, as indicated by a soil test that included both mineralizable N and NO₃-N. The objective of our study was to evaluate the soil test-yield relationship, developed from experimental data at one location in south central Idaho, for predicting N fertilizer needs throughout southern Idaho under various irrigation management practices.

Theory and Basic Relations

Previous investigations showed that N in the soil, measured as NO₃-N and mineralizable N, may not represent all of the N taken up by the sugarbeet crop (5). Although NO₃-N in the sampling zone can be measured, the plants may take up additional NO₃-N from a hardpan

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or from below the sampling depth. Mineralizable N in the sampling zone, as determined in the laboratory, is an index of the supply from this source, but more or less N may be mineralized in the field, and N may be taken up from below the sampling zone. N uptake (Nup) by the crop from various sources is the best measure of N availability and can be expressed as:

$$N_{up} = E_f N_f + \alpha_n N_n + \alpha_m N_m$$
 [1]

where E_f = the efficiency of applied N fertilizer (N_f),

 $\alpha_n = \frac{\text{crop extractable NO}_3-\text{N}}{\text{NO}_3-\text{N in the soil depth sampled}}$

 $N_n = \text{soil NO}_3$ -N in the soil depth sampled,

 $\alpha_{m} = \frac{\text{crop extractable mineralizable N}}{\text{field mineralizable N in soil depth sampled}} \times \frac{\text{field min. N,}}{\text{lab. min. N}}$

 N_m = mineralizable N in the soil depth sampled, as determined by the laboratory mineralization tests.

The more difficult parameters in equation [1] to determine are Ef, α_n , and α_m . Ef can be evaluated by determining total N uptake from about 4 rates of Nf (0, 0.5, 1.0, and 1.5 to 2 times the rate needed for optimum production) or by assuming a value between 0.5 to 0.7 (13). When evaluating Ef, the method and time of fertilizer application normally used by the growers should be used. Determining α_n is more difficult, since $\alpha_m N_m$ is not easily separated from $\alpha_n N_n + \alpha_m N_m$. At least two levels of N_n are needed. These may be approximated for a soil type by applying two excessive N fertilizer treatments for the crop grown before sugarbeets to give two levels of residual Nn. After determining the total Nuptake by sugarbeets on these two treatments, and assuming N_m to be the same on both, α_n and α_m can be approximated by a trial-and-error procedure (assuming a value for $\alpha_{\rm IR}$, calculating $\alpha_{\rm p}$, and solving for $\alpha_{\rm m}$, etc.). The sampling depth for determining N_n and N_m should represent normal sampling depths for the area involved and be consistent with expected future sampling and analyses.

If the entire rooting depth is sampled, and if all of the NO₃-N in this zone is taken up by the crop, α_n will be 1.0. If the entire rooting depth is sampled, but not all of the NO₃-N is taken up, α_n will be less than 1.0. If only part of the rooting depth is sampled and some NO₃-N is taken up from below the sampling depth, α_n will be greater than 1.0.

Previous investigations indicated that when sampling to the cemented zone on a Portneuf silt loam soil in south central Idaho near Twin Falls, $E_f = 0.65$, $\alpha_n = 1.2$, and $\alpha_m = 0.95$ (5). In this study, only E_f could be evaluated because of the lack of different N_n levels.

Previous studies have shown that for maximum sucrose yields, the N requirements per ton of beet roots is 11 ± 1 lbs (5). Less N is required for root production at low levels of available soil and fertilizer N, and

more N is used when the N levels exceed the plant needs for maximum sucrose production.

If 11± 1 lbs of N are needed to produce a ton of fresh roots, then the potential yield, Y, for a sugarbeet field, if limited by N, will be:

$$Y = N_T / (11 \pm 1), N_T / (11 \pm 1) \le Y_F$$
 [2a]

or
$$\frac{Y}{Y_{E}} = \frac{N_{T}}{Y_{E} (11\pm 1)} N_{T} \leq Y_{E} (11\pm 1)$$
 [2b]

where YE is the expected maximum yield under a given management level, when N is not limiting (obtained from farm records), NT is the total "net" N available to the crop (NT = Ef Nf + α_n Nn + α_m Nm). If maximum yields expected from a farmer's management are desired and (α_n Nn + α_m Nm) \leq (11 \pm 1) YE, the N fertilizer needed to make up the deficit, (11 \pm 1) (YE - Y), will be:

$$N_f = \frac{Y_E(11\pm 1) - (\alpha_n N_n + \alpha_m N_m)}{E_f}$$
 [3]

where N_f is the needed N fertilizer, and E_f is the N fertilizer efficiency, expressed as a fraction. The E_f value can be expected to range from 0.5 to 0.7, depending on management practices (13), and was previously found to be 0.65 in this area (5). After harvest, the yield response to N can be evaluated by substituting Y_{max} for Y_E in Equation [2b].

Materials and Methods

During 1971, 32 experiments, involving four N fertilizer treatments, were established throughout southern Idaho. The results from 24 were usable (Table 1). The experimental sites, each 60×100 feet, were located midway between the upper and lower end of irrigated sugarbeet fields. Nutrients, other than N, were applied to the sites at the level used by the farm managers. All other nutrients, except N, were considered adequate from soil and irrigation water sources.

Each experimental site was divided into four $30 - \times 50$ -foot plots, and ammonium nitrate applied at rates 0, 0.5, 1, and 2 times the recommended amount for each site. The recommended amount of N fertilizer was obtained from fertilizer and sugarbeet company fieldmen, based on past fertilizer and cropping histories. In all tests, the recommended N fertilizer rate was applied to the surrounding sugarbeet field by the farm managers. The fertilizer was broadcast and disked into the surface 3 to 4 inches of soil on each site. All cultural operations and irrigations were uniform for the entire field.

Each site was sampled to a depth of 60 inches or to the hardpan in the spring of 1971, before applying fertilizer. Twenty-four cores per

Table 1.—Classification and soil	properties of experimenta	al sites used in the nitrogen study in southern Idaho.

Area	Soil Classification	Root	Previous	Surface soil properties (0-6")		
No.	Designation, Subgroup, and Family	Zone ¹	Crop	pH	O.M.%	N%
3 3 5 5		Southwestern	SEREE		2 7 2	
1	Scism sil, Haplorexollic Durorthid ³	20	Beans	7.80	2.88	0.07
2	Garbutt sil, Typic Torriorthent ⁴	60	Beets	7.80	3.92	0.10
4	Greenleaf sil, Xerollic Haplargid ⁵	30	Onions	7.50	4.23	0.10
6	Elijah sil, Mollic Durargid ⁵	14	Fallow	7.90	3.76	0.10
7	Power sil, Xerollic Haplargid ⁵	30	Sweet Corn	7.80	2.88	0.05
8	sicl	24	Onions	7.20	3.56	0.09
		South Central				
101	Declo sil, Xerollic Calciorthid ⁶	21	Corn	7.90	3.24	0.08
103	Portneuf sil, Xerollic Calciorthid6	24	Potatoes	7.55	0.89	0.08
104	Portneuf sil, Xerollic Calciorthid ⁶	23	Beans	7.65	0.74	0.08
105	Portneuf sil, Xerollic Calciorthid ⁶	24	Potatoes	7.50	1.84	0.0
106	Portneuf sil, Xerollic Calciorthid ⁶	20	Wheat ²	7.55	0.89	0.10
151	Portneuf sil, Xerollic Calciorthid ⁶	17	Potatoes	7.55	1.74	0.10
152	Decker 1, Aquic Calciorthid ⁷	24	Beets	7.50	1.32	0.09
156	Kimama sil, Aridic Calcic Arvixeroll ⁵	24	Wheat	7.75	1.26	0.08
157	Portneuf sil, Xerollic Calciorthid ⁶	18	Wheat	7.60	0.89	0.09
		Southeastern				
201	Portneuf sil, Xerollic Calciorthid ⁶	17	Potatoes	7.95	1.00	0.0
202	Neeley sil, Calciorthidic Haploreroll ³	60	Potatoes	7.45	0.74	0.0
204	Broncho I, Xerollic Camborthid8	16	Beets	7.75	1.05	0.10
205	Portneuf sil, Xerollic Calciorthid ⁶	23	Potatoes	7.60	1.53	0.08
206	Portneuf sil, Xerollic Calciorthid ⁶	18	Potatoes	7.50	0.68	0.0
207	Declo sil, Xerollic Calciorthid ⁶	21	Potatoes	7.35	1.95	0.13
208	Ammon sil, Calciorthidic Haploxeroll9	60	Beets	7.45	1.11	0.10
210	Pancheri sil, Xerollic Calciorthid9	14	Potatoes	7.75	1.53	0.1
211	Bannock 1, Aridic Calcixeroll ¹⁰	23	Beets	7.70	1.32	0.09

¹Soil depth to hardpan or 60 inches

²Straw burned

³Coarse-silty, mixed, mesic

⁴Coarse-silty, mixed, calcareous, mesic

⁵Fine-silty, mixed, mesic

⁶Coarse-loamy, mixed, mesic

⁷Fine-loamy, mixed, mesic

^{*}Sandy-skeletal, mixed, mesic
*Coarse-silty, mixed, frigid
*Oarse-loamy over sand or sandy skeletal, mixed, frigid

site were composited by 6-inch depth increments to the 24-inch depth and by 12-inch increments below that depth. The soil samples were air-dried, ground, and stored until analyzed.

The potential available soil N was determined by extracting NO₃-N from air-dried soil and from 50 g of soil incubated in a 500-ml Erlenmeyer flask for 21 days at 30°C with moisture maintained at approximately 1/3 atm. Moisture loss was minimized by using a one-hole rubber stopper in the flask for aeration during the incubation. The NO₃-N was extracted with a CuSO₄·5H₂O (2.5 g/1) and Ag₂SO₄ (0.167 g/1) solution. The 50-g soil sample was shaken for 10 minutes with 200 ml of extractant, then 1.2 g precipitating mixture, composed of 10 parts of MgCO₃ and 4 parts Ca(OH)₂, were added and the sample shaken again for 5 minutes. Samples were then filtered through Whatman⁴ No.2 filter paper, and an aliquot taken for NO₃-N determination by the phenoldisulfonic acid method essentially as described by Bremner (1).

The difference between the NO₃-N concentrations found in the incubated and air-dried samples was considered the mineralizable N. Small amounts of ammonium-N normally found in these soils were assumed to be oxidized to NO₃-N during incubation and, therefore, were included in the mineralizable N fraction.

Samples of 24 of the youngest fully mature petioles were selected at random from each plot (two in July, two in August, and one in the first part of September). The petioles were cut into ¼-inch sections, dried at 65°C, ground to pass through a 40-mesh sieve, subsampled, and analyzed for NO₃-N, using a nitrate specific ion electrode (11).

The beet tops, crowns, and roots from six uniform 10-foot sections of row were harvested from each treatment at the end of the season to determine root yield, sucrose percentage, sucrose yield, impurity index, and total N uptake. Impurity index (2) and sucrose content were determined on two samples, 30 lbs each, of randomly selected roots from each plot by the Amalgamated Sugar Company, using their standard procedures. The beet pulp (collected during sucrose analysis,) tops, and crowns were dried at 65°C and dry matter determined. The dried samples were ground to pass a 40-mesh sieve, and total N in the samples was determined by the semimicro-Kjeldahl procedure modified to include nitrate (1). Nitrogen uptake was determined by assuming that the percentage N was the same in the fibrous and storage roots, and that the fibrous roots make up 25% of the total harvested root weight (10).

The field numbers, locations, soil classifications, previous crop, and surface soil properties of the 24 experimental sites are given in Table 1. Soil pH was determined using a glass electrode measurement in a soil-water saturated paste, organic matter by a modified method of Walkley and Black (15), and total soil N by the Kjeldahl procedure modified to include nitrates (3).

⁴Mention of trade names or companies is for the benefit of the reader and does not imply endorsement by the U. S. Department of Agriculture.

Results and Discussion

The initial NO₃-N (N_n), mineralizable N (N_m), and total available N (N·r) varied widely between sites and were not consistent within soil series or types (Table 2). A range of 123-236, 127-223, and 104-195 lbs of mineralizable N were released during incubation for the southwestern, south central and southeastern Idaho areas, respectively. The mineralizable N and/or the NO₃-N from below the depth sampled comprised 49 to 81% and averaged 63% of the total N available for plant growth where no N fertilizer was applied. These data indicated that the mineralizable N in the higher organic matter soils of southwestern Idaho could be roughly estimated from their organic matter content, but the lower organic matter soils in south central and southeastern Idaho released 1.5 to 3.0 times more N than would be expected (assuming 5-6% of O.M. is N and 1-2% of this N is released yearly). However, the total N content of the higher and lower organic matter soils were comparable, indicating the N fractions mineralized were similar. Past cropping and management practices over an extended period of years apparently had a pronounced effect on the total N available for plant growth from the mineralization process.

Equation [2a] was evaluated for southern Idaho in 1971 by assuming E_f = 0.65, α_n = 1.2, and α_m = 0.95 (Figure 1). The root yield predicted from N levels generally agreed with yields of the harvested beets. However, on some sites, yields were below those expected, based on available soil N (sites 157, 202), which is apparently due to production difficulties other than those under study (Figure 1a). Yields on four of the sites were higher than those expected from the available soil N (6, 7, 101, 201). The sugarbeet plant becomes much more efficient in the use of N for root production at lower levels of available N, as shown by sites 101 and 201. Yields on both of these sites were predictable from soil N levels, after adequate N fertilizer was applied to get maximum yields (Figure 1b). Site 6 had been fallowed for 2 years before this study and had accumulated large amounts of NO₃-N just above or in the hardpan. The assumed α_n value of 1.2 probably was too low, and yields indicated that α_n should have been near 1.6 for this site: Yields from the check plot on site 7 cannot be explained from either the amount of N available from soil sources or from the amount of N taken up by the plants per ton of beet roots (Table 2). The sugarbeet plants apparently used the soil N much more efficiently on this site. Yield was predictable, however, when adequate N was present for maximum root and sucrose yield.

If the root yield potential for any sugarbeet field is known and other factors are not limiting, then the amount of N fertilizer necessary for maximum yields can be predicted, using equation [3] as shown in Table 2. The predicted N required for site 6 exceeded the N necessary for maximum sucrose yields enough to significantly reduce the sucrose percentage and sucrose yield. Fertilizer predictions based on soil test $\alpha_{\rm D} N_{\rm D} + \alpha_{\rm D} N_{\rm D}$ were superior in southern Idaho to procedures used

Exp.				Yi	Yield N Fertilizer Needed				N uptake		NT'				
Area		Soil		Mea	sured	-	Recomn	endation	ıs ba	sed on	Max.	Check	Max.	Check	Max.
No.	Nn	N _m	NT'	Y min.	Y max.	NT'	NO ₃	N ² NO ₃	N×	1.47 ³ Fieldmen	Yield	plot	Yield	plot	Yield
		lbs N/A		1	.1			llis	1. 1				llss."	S/ [
1	101	171	3494	28.3	28.3	()-()	81		41	90	()	11.2	11.2	12.3	12.3
2	81	236	321	20.4	20.1	()-()	107		116	150	()	8.3	8.3	15.7	15.7
4	165	210	398	28.4	28.4	0.0	118		185	200	0	11.0	1.1.0	14.0	14.0
6	101	1.53	267	28.5	28.5	28 115	183		175	180	EE.	10.0	10.0	9.4	9.1
7	96	123	232	26.3	30 1	111-205	207		225	150	150	1.9	× 7	8.8	10.9
8	135	148	303	25.5	29.9	0.86	163		225	230	115	9.0	10.2	11.9	12.6
101	11	132	175	20.6	27.4	152 237	232		242	160	160	5.4	9.3	8.5	10.2
103	79	201	286	25.5	25.5	0-31	175		203	1.10	0	6.3	6.3	11.2	11.2
104	79	159	2761	24.7	25.8	()-52	118		160	175	57	6.1	9.3	11.2	12.9
105	104	127	245	20.1	20.1	()-()	96		140	150	()	9.6	9.6	12.2	12.2
106	77	187	270	24.3	26.7	0.77	189		216	100	50	8.1	8.1	11.1	11.3
151	130	189	336	22.8	22.8	0.0	97		121	150	11	11.0	11.0	14.7	14.7
152	122	223	358	21.2	21.2	()-()	89		102	160	()	12.1	12.1	16.9	16.9
156	3.4	111	178	14.1	111	()-()	106		115	200	()	5.8	5.8	12.6	12.6
157	88	157	255	18.3	20.8	()-()	119		116	150	7.5	11.8	11.9	13.9	14.6
201	62	101	1.73	19.3	22.0	72-140	1.57		170	120	120	8.8	114	9.0	11.3
202	134	129	283	20.9	25.1	0.28	86		178	120	60	6.2	10.9	13.5	12.8
204	50	128	182	177	17.7	01-46	126		130	130	11	12.1	12.1	10.3	10.3
205	116	1.1.1	273	22.4	26.7	0.72	150		206	180	90	11.1	12.7	12.2	12.4
206	117	116	251	21.6	23.8	0-51	120		159	120	titt	12.0	13.5	11.6	12.2
207	171	168	365	25.7	25.7	(1-()	85		59	100	0	10.7	10.7	14.2	11.2
208	1-15	195	359	28.7	28.7	0.0	69		207	130	0	10.0	10.0	12.5	12.5
210	140	194	352	23.5	23.5	()-()	94		71	1:30	0	12.0	12.0	15.0	15.0
211	.5()	183	234	21.0	21.0	0-28	159		156	120	()	7.8	7.8	11.1	11.1
Avg.	101	163	280	22.9	24.4	15-19	132		155	117	10	9.3	10.2	12.2	12.6

Total available soil and fertilizer $N(\alpha_n/N_n)^2/\alpha_m/N_m + E_f/N_f)$ in the top 36 inches or to the hardpan.

²Calculated N fertilizer needs from residual NO₁ N to the 60-inch soil depth or hardpan by methods of Reuss and Rao (12).

³Calculated N tertilizer needs from residual NOx. N in the top 12 inches × 1.47 by methods of Reuss and Rao (12)

⁴N tertilizer applied to experimental area by farm manager.

^{*}Calculated N fertilizer need for maximum yield (Ymax) if 10 to 12 lbs N/1 of beet roots is required

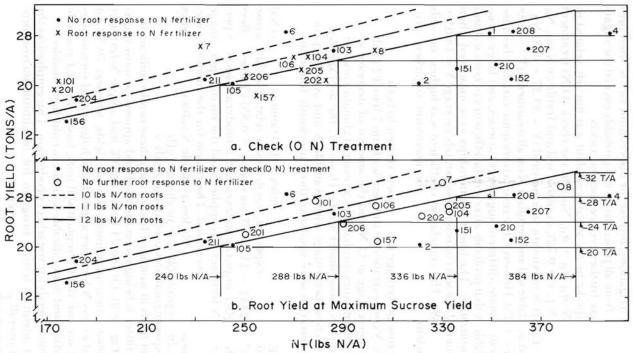


Figure 1a and 1b.—Effect of available N on the root yield of sugarbeet. (Root yields should be near the 10 to 12 lbs N/ton of roots lines until sufficient N is available for maximum root yield. Excess N for the root yield potential of the sugarbeet field will maintain root yield while increasing the total available soil and fertilizer N (N_T) per ton of roots. Solid vertical and horizontal lines represent the N level necessary to produce a given yield level if 12 lbs of N is required to produce a ton of beet roots.)

by fieldmen based on past fertilization and cropping histories and predictions based only on available NO₃-N (Table 2). However, if cultural practices, irrigations, disease, or insects limit yields, then predicted required N, from all procedures, will probably exceed the sugarbeet needs and may further reduce sucrose yields.

The total N uptake by the sugarbeet plant was linearly related to the available soil N (α_n N_n + α_m N_m) on the check (O N) treatment ($\hat{Y} = 8.60 + 0.73$ N_T, r = 0.72), available soil and fertilizer N (α_n N_n + α_m N_m + E_f N_f) at maximum sucrose yield ($\hat{Y} = 33.37 + 0.70$ N_T, r = 0.67), and available soil and fertilizer N on all treatments ($\hat{Y} = 47.88 + 0.67$ N_T, r = 0.75). However, the results were quite variable, because of variations in climate, different sugarbeet varieties, and large variations in available N that probably affected the efficiency of both soil and fertilizer N utilization. The linear relationships were improved when each experimental site was considered separately (average $\hat{Y} = 18.03 + 0.78$ N_T, r = 0.88). The correlation coefficient was highest on soils initially deficient in available soil N for maximum sugarbeet production.

The sucrose percentage was inversely related to both N_T and the average NO₃-N concentration in the petioles from 7/6 to 9/2 (Table 3). The sucrose percentage at O N and the amount of decrease with N fertilizer varied with sites, probably as a result of climatic conditions or factors other than those under study. However, the average rate of decrease in sucrose percentage with available N and average petiole NO₃-N concentration were similar to those previously found in a detailed study at one site (6).

The impurity index (Impurity Index = [10 (amino N) + 3.5 (Na) + 2.5 (K)]/sucrose %), as expected, was inversely related to the sucrose percentage ($\hat{Y} = 2441 - 115 \times \%$ sucrose, r = 0.85) (Table 3). Beet roots with the higher impurity indexes had a moderate sucrose percentage when no N fertilizer was applied, and the sucrose percentage decreased as N fertilizer increased. The reason for the moderate sucrose percentage at comparatively low available N values at certain sites is unknown, but is probably due to climatic effects.

If the experimental sites used in this study were representative of the sugarbeet fields in southern Idaho, then these results indicate that excess N fertilizer is being recommended and applied on most sugarbeet fields (Figure 2). Of the 24 fields, 29.5% had ample N fertilizer applied to obtain near maximum sucrose yields, whereas 70.5% of the fields had excess N fertilizer applied which decreased sucrose percentage and sucrose yield. Preplant soil tests, based on total available N from residual and mineralizable sources, would have enabled predicting optimum needs for sugarbeet production. This emphasizes the need for an adequate testing program for maximum sucrose production and overall profits. However, as long as the payment to an individual grower is based on root tonnage and average sucrose percentage for the district, controlling N fertilizer to maximize sucrose

Table 3Effect of available N and average petiole NO3-N o	on the sucrose percentage and impurity index of sugarbeets.
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Area No.	% Sucrose		Imp. Index		Regression Analysis					
	Check	2 × Rec.	Check	2 × Rec.	Available N(N _T) ¹	r	Avg. petiole NO ₃ -N ²	r		
1	17.0	16.0	526	710	$\hat{Y} = 18.8 - 4.1 \times 10^{-3} \text{ X}$	-0.67	$\hat{Y} = 19.5 - 2.1 \times 10^{-4} \text{ X}$	-0.71		
2	17.0	14.8	319	644	$\hat{Y} = 19.8 - 9.7 \times 10^{-3} \text{ X}$	-0.89	$\hat{Y} = 17.3 - 1.8 \times 10^{-4} \text{ X}$	-0.88		
4	15.0	13.5	496	934	$\hat{Y} = 17.1 - 5.7 \times 10^{-3} \text{ X}$	-0.95	$\hat{Y} = -15.7 - 1.3 \times -10^{-4} \text{ N}$	-0.92		
6	16.8	14.8	459	875	$\hat{Y} = 17.6 - 5.9 \times 10^{-3} \text{ X}$	-0.57	$\hat{Y} = 18.8 - 3.4 \times 10^{-4} \text{ X}$	-0.94		
7	17.7	16.5	293	382	$\hat{Y} = 19.4 - 6.5 \times 10^{-3} \text{ X}$	-0.97	$\hat{Y} = 18.0 - 2.3 \times 10^{-4} \text{ X}$	-0.96		
8	16.4	12.4	436	1,069	$\hat{Y} = 20.9 - 14.4 \times 10^{-3} \text{ X}$	-0.95	$\hat{Y} = 19.2 - 3.7 \times 10^{-4} \text{ X}$	-0.96		
101	17.9	16.2	298	577	$\hat{Y} = 19.9 - 9.1 \times 10^{-3} \text{ X}$	-0.91	$\hat{Y} = 18.4 - 1.0 \times 10^{-4} \text{ X}$	-0.93		
103	18.0	16.2	378	670	$\hat{Y} = 21.4 - 10.8 \times 10^{-3} \text{ X}$	-0.94	$\hat{Y} = 18.9 - 1.3 \times 10^{-4} \text{ Y}$	-0.92		
104	18.4	16.3	275	599	$\hat{Y} = 21.3 - 10.0 \times 10^{-3} \text{ X}$	-0.95	$\hat{Y} = 18.9 - 1.6 \times 10^{-4} \text{ X}$	-0.95		
105	18.7	16.3	430	725	$\hat{Y} = 21.5 - 11.8 \times 10^{-3} \text{ X}$	- 0.99	$\hat{Y} = 20.2 - 1.6 \times 10^{-4} \text{ X}$	-0.98		
106	17.8	17.1	267	442	$\hat{Y} = 19.4 - 5.9 \times 10^{-3} \text{ X}$	-0.93	$\hat{Y} = 18.0 - 0.9 \times 10^{-4} \text{ X}$	-0.99		
151	.16.1	14.4	645	721	$\hat{Y} = 19.1 - 8.9 \times 10^{-3} \text{ X}$	-0.99	$\hat{Y} = 17.4 - 1.5 \times 10^{-4} \text{ X}$	-0.84		
152	16.0	15.6	603	731	$\hat{Y} = 16.4 - 1.8 \times 10^{-3} \text{ X}$	-0.44	$\hat{Y} = 17.0 - 0.9 \times 10^{-4} \text{ X}$	-0.52		
156	18.9	16.6	402	603	$\hat{Y} = 21.3 - 10.4 \times 10^{-3} \text{ X}$	- 0.89	$\hat{Y} = 19.9 - 1.4 \times 10^{-4} \text{ X}$	-0.80		
157	16.1	13.2	737	1,159	$\hat{Y} = 19.5 - 14.8 \times 10^{-3} \text{ X}$	- 0.93	$\hat{Y} = 24.2 - 5.4 \times 10^{-4} \text{ X}$	-0.99		
201	17.1	16.8	491	602	$\hat{Y} = 17.5 - 2.3 \times 10^{-3} \text{ X}$	- 0.64	$\hat{Y} = 17.2 - 0.5 \times 10^{-4} \text{ X}$	-0.67		
202	18.9	16.9	-		$\hat{Y} = 21.4 - 11.7 \times 10^{-3} \text{ X}$	- 0.66	$\hat{Y} = 19.7 - 2.1 \times 10^{-4} \text{ X}$	-0.95		
204	18.0	15.5	-		$\hat{Y} = 21.1 - 15.6 \times 10^{-3} \text{ X}$	- 0.98	$\hat{Y} = 22.4 - 3.4 \times 10^{-4} \text{ X}$	-0.99		
205	15.7	15.2	_		$\hat{Y} = 16.1 - 2.1 \times 10^{-3} \text{ X}$	-0.70	$\hat{Y} = 16.5 - 0.9 \times 10^{-4} \text{ X}$	-0.82		
206	17.4	15.1			$\hat{Y} = 20.2 - 13.1 \times 10^{-3} \text{ X}$	-0.89	$\hat{Y} = 22.0 - 3.9 \times 10^{-4} \text{ X}$	-0.89		
207	15.9	16.1	-	_	$\hat{Y} = 14.5 + 2.7 \times 10^{-3} \text{ X}$	+ 0.39	$\hat{Y} = 16.8 - 1.0 \times 10^{-4} \text{ X}$	-0.37		
208	16.3	15.6	-		$\hat{Y} = 17.2 - 3.7 \times 10^{-3} \text{ X}$	- 0.53	$\hat{Y} = 19.2 - 2.5 \times 10^{-4} \text{ X}$	-0.99		
210	17.4	14.9	_		$\hat{Y} = 23.0 - 15.9 \times 10^{-3} \text{ X}$	_ 0.95	$\hat{Y} = 23.8 - 4.4 \times 10^{-4} \text{ X}$	-0.93		
211	17.9	15.7	1-	나라는 본트	$\hat{Y} = 21.3 - 14.4 \times 10^{-3} \text{ X}$	_ 0.99	$\hat{Y} = 17.4 - 1.5 \times 10^{-4} \text{ X}$	-0.84		
Avg.	17.2	15.5	441	* 715	$\hat{Y} = 19.4 - 8.6 \times 10^{-3} \text{ X}$	- 0.82	$\hat{Y} = 19.0 - 2.1 \times 10^{-4} \text{ X}$	-0.86		
	For all poi	nts			$\hat{Y} = 19.8 - 9.4 \times 10^{-3} \text{ X}$	- 0.69	$\hat{Y} = 18.0 - 1.5 \times 10^{-4} \text{ X}$	-0.57		

 $^{{}^1\}hat{Y}=\%$ sucrose, X= available $N(N_T)$ ${}^2\hat{Y}=\%$ sucrose, X= average petiole NO_3-N concentration from 7/6 to 9/2.

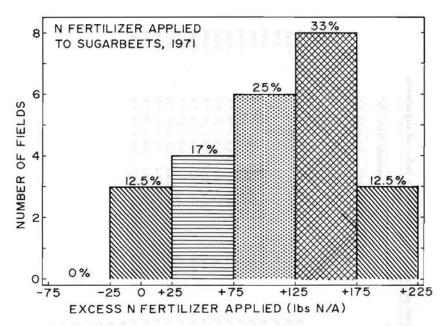


Figure 2.—Distribution of excess N fertilizer recommended and applied to sugarbeet fields in southern Idaho in 1971.

production has no economic advantage over maximizing yield of beets per acre.

In conclusion, the NO₃-N level in the soil has been shown in these and other studies (5, 8, 12) to be an excellent indicator of the N fertilizer needs of sugariseets for maximum sucrose production, provided that the mineralization capacity of the soil and the yield potential of the field are known. The amount of N supplied from mineralizable sources in a uniformly cropped and fertilized field is expected to remain reasonably constant from one year to the next, if adequate but not excess N fertilizer is supplied yearly for the crop grown (5). Once the mineralization capacity of a soil has been determined, this test need not be repeated yearly. Soil sampling and laboratory analyses, to determine the amount of NO₃-N in the rooting zone, when combined with the predetermined mineralizable N, would enable accurate N fertilizer recommendations. Mineralizable N should be redetermined every few years, particularly following forage legumes or unusual fertilizer practices.

The use of this soil test should indicate the optimum N levels for maximum refined sucrose production, provided that proper irrigation levels are used (9). Excessive irrigations, particularly early in the season before the period of maximum N uptake by the crop, will move part of the N supply out of the root zone and make it unavailable to the sugar-

beets. Although the testing for and the application of proper amounts of N fertilizer before the period of maximum plant N uptake are important, a mid-season verification of the N status of the crop by petiole analysis (4) would help to determine the accuracy of N fertilizer application recommendations in relation to the irrigation levels, and should permit fertilizer and irrigation adjustments during the current and future years for maximum refined sucrose production.

Summary

Sugarbeets (*Beta vulgaris* L.) were grown at four N fertilizer levels on 24 sites throughout southern Idaho to determine root yield, sucrose percentage, sucrose yield, impurity index, and plant N uptake in relation to the residual, mineralizable, fertilizer N, and petiole NO₃-N. A soil test to measure both mineralizable and nitrate-nitrogen level of a soil serves as a valuable guide in recommending nitrogen fertilizer for sugarbeets over a wide area of southern Idaho. The use of this test will enable the optimum application of nitrogen fertilizer before planting or side-dressing early in the season, before the period of highest nitrogen uptake by the plant, to obtain maximum refinable sucrose production and profits to both the producer, when paid on a refined sucrose basis, and processor.

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Literature Cited

 Bremner, J. M. 1965. Inorganic forms of nitrogen. In C. A. Black (ed.) Methods of Soil Analysis. Part 2. Agronomy 9:1179-1237. Amer. Soc. of Agron., Madison, Wis.

(2) CARRUTHERS, A., J. F. T. OLDFIELD, and H. J. TEAGUE. 1962. Assessment of beet quality. Fifteenth Annual Technical Conference of the British

Sugar Corporation, Limited, Nottingham, England.

(3) CARTER, J. N., O. L. BENNETT, and R. W. PEARSON. 1967. Recovery of fertilizer nitrogen under field conditions using Nitrogen—15. Soil Sci. Soc. Amer. Proc. 31:50-56.

(4) CARTER, J. N., M. E. JENSEN, and S. M. BOSMA. 1971. Interpreting the rate of change in nitrate-nitrogen in sugarbeet petioles. Agron. J. 63:669-674.

(5) CARTER, J. N., M. E. JENSEN, and S. M. BOSMA. 1974. Determining nitrogen fertilizer needs for sugarbeets from residual soil nitrate and mineralizable nitrogen. Agron. J. 66:319-323.

(6) CARTER, J. N., M. E. JENSEN, B. J. RUFFING, S. M. BOSMA, and A. W. RICHARDS. 1972. Effect of nitrogen and irrigation on sugarbeet production in southern Idaho. J. Am. Soc. Sugar Beet Technol. 17: 5-14.

(7) HILLS, F. JACKSON and ALBERT ULRICH. 1971. Nitrogen nutrition. p. 112-135. In R. T. Johnson, J. T. Alexander, G. E. Rush, and G. R. Hawkes (eds.) Advances in Sugarbeet Production: Principles and Practices. The Iowa State University Press, Ames, Iowa.

(8) JAMES, D. W., A. W. RICHARDS, W. H. WEAVER, and R. L. REEDER. 1971. Residual soil nitrate measurements as a basis for managing nitrogen fertilizer practices for sugarbeets. J. Am. Soc. Sugar Beet Technol. 16:313-322.

(9) JENSEN, M. E. and L. J. ERIE. 1971. Irrigation and water management, p. 189-222. In R. T. Johnson, J. T. Alexander, G. E. Rush, and G. R. Hawkes (eds.) Advances in Sugarbeet Production: Principles and Practices. The Iowa State University Press, Ames, Iowa.

(10) KELLEY, J. D. and A. ULRICH. 1966. Distribution of nitrate nitrogen in the blades and petioles of sugarbeets grown at deficient and sufficient levels of nitrogen. J. Am. Soc. Sugar Beet Technol. 14:106-116.

(11) MILHAM, P. J., A. S. AWAD, R. E. PAULL, and J. H. BULL. 1970. Analysis of plant, soils and waters for nitrate by using an ion-selective electrode. Analyst 95:751-757.

(12) Reuss, J. O. and P. S. C. Rao. 1971. Soil nitrate nitrogen levels as an index of nitrogen fertilizer needs of sugarbeets. J. Am. Soc. Sugar Beet Technol. 16:461-470.

(13) STANFORD, GEORGE and A. S. HUNTER. 1973. Nitrogen requirements of winter wheat (*Triticum aestivum*, L.) varieties 'Blueboy' and 'Redcoat'. Agron. J. 65:442-447.

(14) STANFORD, GEORGE and S. J. SMITH. 1972. Nitrogen mineralization potentials of soils. Soil Sci. Soc. Amer. Proc. 36:465-472.

(15) WALKLEY, A. and I. A. BLACK. 1934. An examination of the Degtjareff method for determining soil organic matter, and a proposed modification of the chromic acid titration method. Soil Sci. 37:29-38.